

Response of Cut Carnation cv. Tempo to Essential Oils and Antimicrobial Compounds

Davood Hashemabadi

Department of Horticultural Science
Rasht Branch, Islamic Azad University, Rasht, Iran

Abstract: Carnation cut flower is sensitive to ethylene, which its vase life reduces depending on ethylene production and vascular occlusion. Like other cut flowers, carnation is sensitive to bacterial infection on stem end, bacterial contamination produces toxic compounds and it affects ethylene production that ultimately shortens vase life. In order to study on effect of essential oils, 8-hydroxyquinoline sulphate and aluminum sulphate on vase life and postharvest characteristics of cut carnation (*Dianthus caryophyllus* L. cv. Tempo), a RCD experiment with 13 treatments: Artemisia oil at 3 levels (4, 8 and 12%), Anethum oil at 3 levels (4, 8 and 12%), 8-hydroxyquinoline sulphate at 3 levels (200, 400 and 600 mg l⁻¹) and aluminum sulphate at 3 levels (100, 200 and 300 mg l⁻¹), 3 replications, 39 plots and 156 cut flowers was carried out. The measured traits were stem end bacterial colonies population, ethylene production, water uptake, fresh weight loss and vase life. The results showed that 100 mg l⁻¹ aluminum sulphate, 200 mg l⁻¹ 8-hydroxyquinoline sulphate, 12 % Artemisia and Anethum oils were the best treatments and enhanced vase life & qualitative features.

Keywords: Carnation, Artemisia oil, Anethum oil, Aluminium sulphate, 8-hydroxyquinoline sulphate, vase life, number of bacteria in vase solution, petal protein.

1. INTRODUCTION

Carnation (*Dianthus caryophyllus* L.) is native to the Mediterranean region to Central Asia, belongs Caryophyllaceae family that has long been under cultivation [25]. Carnation is among the world's top three cut flowers. The two species *Dianthus barbatus* & *Dianthus caryophyllus* are cultivated as commercial varieties [13]. Carnation cut flowers are so sensitive to ethylene that causes senescence and vase life reduction [19]. Carnation is also susceptible to the bacterial contamination on stem end, that produces toxic compounds and affects ethylene production which reduces vase life which has petal's enrolling as its most obvious sign of senescence [4-31]. Given the publicity of this problem among cut flowers, it seems to be necessary to find a solution. Essential oils act as incorporating environmentally friendly and highly effective antimicrobial compounds [9-27]. Herbal essential oils are natural compounds that are known as secondary metabolites which have powerful effect on pathogens control and their antimicrobial impact is proven as well that increases the vase life of cut flowers and their use is becoming widespread recently [39-41]. Artemisia (*Artemisia*

dracunculus) belongs to Asteraceae family that has lots of medical properties, Dill (*Anethum graveolens*) is an annual or biennial plant that sometimes reaches its height to one meter [34]. Mousavi Bazaz & Tehranifar [32] investigated the effects of cumin, mint and thyme extract (50 and 100 mg l⁻¹) and ethanol (4 & 7%) on the vase life of cut *Alstroemeria* (*Alstroemeria* sp.) and concluded that all treatments had a positive effect on vase life and water uptake and also 50 mg l⁻¹ thyme extract extended vase life about 2 days more than the control. Solgi et al. [40] studied on the effect of different herbal essential oils including Carvacrol, Thyme oil and Zataria essential oil & silver nanoparticles on the vase life of gerbera (*Gerbera jamesonii* cv. 'Dune'), and reported that treatment with 50 and 100 mg l⁻¹ carvacrol increased the vase life of about 8 days as compared to the control. 8-hydroxyquinoline sulfate is used mainly to acidification of the solution and combined with divalent ions such as iron and copper to prevent vascular occlusion, that can prevent enzymatic activities which lead to vascular blockage [9-21]. Tar & Hassan [42] evaluated the effect of various levels of 8-hydroxyquinoline sulfate on the postharvest life of *Aster* sp. cut flower and concluded that the use of 400 mg l⁻¹ of this compound increases the vase life. It is proved that using calcium, aluminum, boron, nickel & zinc salts can extend vase life of cut flowers [9].

In addition to the inhibitory effect of aluminum sulfate on microorganisms activity, it reduces the transpiration of cut flowers. It appears considering the carnation leaves shape and cuticle thickness, the effects of aluminum sulfate on reduction of transpiration and stomatal exchange rate is less. Previous studies show that the use of aluminum sulfate, especially at high concentrations may cause damage to the leaves of some flowers such as roses (*Rosa hybrida* L.) [10]. Liao et al. [30] investigated the effect of aluminum sulfate at different concentrations (50, 100 & 150 mg l⁻¹) on the vase life of cut *Eustoma grandiflora* and concluded that using 150 mg l⁻¹ increased vase life up to 15.4 days. Additionally, aluminum sulfate enhanced water uptake and fresh weight. Hojati et al. [20] investigated different chemicals like cobalt chloride and aluminum sulfate on *Eustoma grandiflora* vase life and resulted that aluminum sulfate extends vase life of cut flowers. This study is about comparison the effect of different

concentrations of herbal essential oils and other antimicrobial compounds on the vase life of cut carnation cv. 'Tempo'.

2. MATERIALS AND METHODS

Cut carnation (*Dianthus caryophyllus* L. Cv. 'Tempo') provided from a commercial manufacturer's and were equalized at 50 cm height, then they were re-cut from stem end; it's because of vascular blockage inhibition. Then they immediately transferred to the postharvest laboratory with the following condition: $12 \mu\text{Ms}^{-1}\text{m}^{-2}$ light intensity, $20 \pm 2^\circ \text{C}$ temperature, 12 hours day length and a relative humidity of 60 to 70 percent to traits assessment. 5 cutflowers were placed in each plastic 2 lit volume vase and then were treated with specific concentrations of antimicrobial compounds.

This experiment is based on a completely randomized design with 13 treatments (Artemisia essential oil at 3 levels (4, 8 and 12%); 8-hydroxyquinoline sulfate at 3 levels (200, 400 & 600 mg l^{-1}) and aluminum sulfate at 3 levels (100, 200 and 300 mg l^{-1}) along with control plots was carried out in 3 replicates and 39 plots. The pulse treatment completed after 24 hours then flowers were transferred into other vases containing 500 ml 3% sucrose + 200 mg l^{-1} 8-hydroxyquinoline sulfate preservative solution.

The measured traits were vase life, fresh weight loss, water uptake, ethylene production and bacterial populations on the stem end. Vase life is characterized with leaf wilting index or leaf senescence and flowers wilting (fig. 1) [33]. To evaluate the fresh weight loss, flowers fresh weight were measured at the first & the last day of vase life, and also considering the re-cut part weight, their difference were recorded. Due to the preservative solution initial content (500 ml), the absorption was measured according to each day reduction (both evaporation & stem uptake), the solution absorption was calculated & has been divided to the initial flowers weight. To measure the amount of ethylene released from each pot, a flower was selected and after cut to 25 cm was weighted & then transferred into smaller pots containing 30 ml 8-hydroxyquinoline citrate 250 mg l^{-1} & each small pot was placed inside jars and sealed completely.

In order to sample the air inside jars, wooden doors of each jar was equipped with a septum. Gas samples were taken to Tehran University Gas Analysis Laboratory. The released ethylene produced with the flower were measured with C-8 AIT measuring device Shimadzu model. For stem end bacterial population counting, about 2 cm (0.5 g) were cut from the stem end 24 hours after treatment with antimicrobial compounds; also for preservative solution bacterial population counting, a 2 ml sample were isolated from each pot and the evaluation were performed according to Liu et al. [30] method.

3. RESULTS AND DISCUSSION

3.1. Vase life

Analysis of variance showed that the effect of different treatments on vase life was statistically significant at 5% level. Mean comparisons revealed that 100 mg l^{-1} aluminum sulfate, 200 mg l^{-1} 8-hydroxyquinoline sulfate, 12% Artemisia & Tarragon essential oils were the most effective treatments with 17.9, 16.7, 16.1 & 15.1 days vase life respectively (Tab. 1, Fig. 2). On present study, vase life enhancement can be described with antimicrobial properties of the mentioned compounds that water absorption improvement with vascular blockage prevention so it delays water deficiency related wilting [8-22]. Jalili Marandi et al. [22] investigated the effects of salicylic acid, herbal essential oils and silver thiosulfate (STS) on the vase life of cut roses and found that *Carum copticum* essential oil at the concentration of 500 mg l^{-1} significantly (about 2 days) improved the vase life as compared to the control. Mousavi Bazaz & Tehranifar [32] on their study on *Alstroemeria* & herbal essential oils, ethanol and methanol impact, found that treatment with 50 & 100 mg l^{-1} had the best effect on water absorption and vase life. Solgi et al. [40] studied different compounds effect on cut gerbera cv. 'Dune' and resulted that 50 & 100 mg l^{-1} caused a two-fold increase on vase life as compared to the control. Faraji [14] found that 8-hydroxyquinoline citrate & aluminum sulfate had the best impact on vase life extend, flower diameter, opening percent and flower quality of cut rose cv. 'Marosia'.

Liao et al. [29] examined the effects of aluminum sulfate at concentrations of 50, 100, and 150 mg l^{-1} on cut *Lisianthus (Eustoma grandiflora)* and concluded that 150 mg l^{-1} of that extends vase life up to 15.4 days, additionally aluminum sulfate enhanced water uptake and fresh weight. Hojjati et al. [20] investigated the effect of different chemical treatments like cobalt chloride and aluminum sulfate on vase life of cut *Lisianthus* and found that aluminum sulfate can increase vase life. Khan et al. [26] studied the effect of aluminum sulfate and sucrose on the properties tulip (*Tulipa hybrida*) properties and concluded that aluminum sulfate increases the relative water content in leaves and petals 64.5% & 58.7% respectively. Anju et al. [2] examined the effect of 8-hydroxyquinoline citrate and sucrose on the vase life of cut chrysanthemum (*Chrysanthemum morifolium* L.) and found that these compounds caused an increase in fresh weight and vase life as compared to the control. Tar & Hassan [42] investigated the impact of various levels of 8-hydroxyquinoline sulfate on (*Aster* sp.) vase life and concluded that the use of 400 mg l^{-1} of this compound increases vase life.

3.2. Fresh weight loss

Analysis of variance showed that the effects of different treatments on fresh weight reduction was significant at the 5% level. Mean comparisons also revealed that 100 mg l^{-1}

aluminum sulfate, 200 mg^l⁻¹ 8-hydroxyquinoline sulphate and 12% Artemisia & Dill essential oil were the most effective treatments with 2.89, 4.2, 4.4 & 4.3 grams loss respectively (Fig. 3, Table 1). Water uptake and water relations improvement with antimicrobial compounds prevent fresh weight loss [6- 15-43]. Liao et al. [30] on their investigation on cut Lisiyanthus and applying different aluminum sulfate concentrations found that aluminum sulfate enhanced fresh weight as compared to the control. These researchers also described this enhancement with a better corolla outreach. Hydroxyquinoline enhanced cut roses fresh weight and vase life significantly [21]. These results are in accordance with [11-3-28].

3.3. Water absorption

The analysis of variance showed that the effects of different treatments on water absorption were significant at 5% level. Mean comparison showed that among all treatments, 100 mg^l⁻¹ aluminum sulfate, 200 mg^l⁻¹ 8-hydroxyquinoline sulfate and 12% Artemisia & Dill essential oil with 1.96, 1.94, 1.80 & 1.78 ml g⁻¹ F.W. gave the best results (Fig. 4, Tab. 1). Improving water relations and hydraulic conductivity in cut flower arises from vascular obstruction prevention which ultimately will improve water absorption [16]. Another reason for superiority of mentioned compounds is microorganisms activity control (bacteria & fungi) that prevents vascular obstruction. Our results match with [22-7]. Nabigol et al. [33] found that antiseptic and anti ethylene compounds and also antibiotics increase water absorption significantly. Anju et al. [2] studied the effect of hydroxyquinoline and sucrose on vase life of chrysanthemum (*Chrysanthemum morifolium* L.) and concluded that these compounds caused an increase in fresh weight and vase life and improved quality in comparison with the control. Khan et al. [26] investigated the impact of aluminum sulfate and sucrose on the properties of tulip (*Tulipa hybrida*) and resulted that aluminum sulfate increases relative water content in leaves and petals 64.5% and 58.7% respectively.

3.4. Bacterial population on stem end

Analysis of variance showed that the effects of different treatments on stem end bacterial population were significant at 1% level. Mean comparison revealed that among all treatments, 100 mg^l⁻¹ aluminum sulfate, 200 mg^l⁻¹ 8-hydroxyquinoline sulfate and 12% Artemisia & Dill essential oil were the most effective treatments that have 14, 25, 29 & 30.4 log₁₀ CFU ml⁻¹ population respectively that have a reduction as compared to the control stem ends bacterial contamination (Fig. 5, Tab. 1). The effect of antimicrobial compounds used in these experiments is defined by disrupting the pathogens function & the respiratory chain which prevents pathogens activity and ultimately causes death (18-40). Esfandiari et al. [12] on their experiment on cut lily (*Lilium longiflorum* cv. 'Shocking') found that antimicrobial compounds extend vase life by interfering bacterial cell

division, these results are in accordance with Liu et al. [30] & Shanani [38] about positive impact of antimicrobial compounds and herbal essential oils on pathogenic contamination control and vase life improvement. Kazemi and Ameri [23] investigated the effects of herbal essential oils, silver nanoparticles and salicylic acid on bacterial contamination of carnation and found that all treatments significantly reduced bacterial populations much more than the control. Kazemi et al. [24] showed that the use of vase life extending compounds at 1.5 & 3 mM concentrations decreased carnation cut flower bacterial population significantly (about 30-35%). Anju et al. [2] examined the effects of hydroxyquinoline and sucrose on vase life of cut chrysanthemum (*Chrysanthemum morifolium* L.) and concluded that these compounds enhance fresh weight and improve vase life and quality traits in comparison with the control. Oraee et al. [35] investigated the effect of different levels of silver nanoparticles and Thyme essential oil on vase life and bacterial populations on stem end & in preservative solution of cut gerbera, they found that all these compounds had a positive effect in controlling bacterial contaminations, they reported that 100 mg^l⁻¹ Thyme oil and 4 mg^l⁻¹ silver nanoparticles with two-fold increase in vase life were the most effective treatments. Hojjati et al. [20] evaluated the effects of different chemical treatments such as cobalt chloride and aluminum sulfate on vase life of cut Lisiyanthus and reported that aluminum sulfate treatment increased the vase life and improved postharvest quality. Elgimabi and Ahmed [11] studied on the effects of 8-hydroxyquinoline sulfate and sucrose on cut roses vase life and found that 100 mg^l⁻¹ 8-hydroxyquinoline sulfate caused doubled vase life for treated flowers as compared to the control and also enhanced other vase life related indexes.

3.5. Ethylene production

Analysis of variance showed that the effects of different treatments on ethylene production were significant at 5% level. Mean comparison revealed that among all treatments and 100 mg^l⁻¹ aluminum sulfate, 200 mg^l⁻¹ 8-hydroxyquinoline were the most effective treatments with 0.201 & 0.214 nl⁻¹ per hour per gram fresh weight significantly less ethylene concentration respectively (Fig. 6, Tab. 1). The impact of these antimicrobial compounds on cut carnation quality enhancement is obvious given the role of antimicrobial compounds on the uptake and transport of nutrients, reduced respiratory rate and reduced the amount of ethylene. Our findings in reducing ethylene production matches with [17]. Reid et al. [36] found that the use of STS on cut carnation cv. "White Seam" in the 1 to 4 molar ratio between silver and thiosulfate was satisfactory. Anti ethylene and antimicrobial compounds due to their stem end bacterial contamination control, can stimulate ethylene production indirectly, can control ethylene production and extend vase life of cut carnation and gerbera (4-5-28). The results of mentioned

studies about ethylene production control is consistent with the results of the current study.



Figure 1. Cut carnation cv. 'Tempo' at the last day of vase life

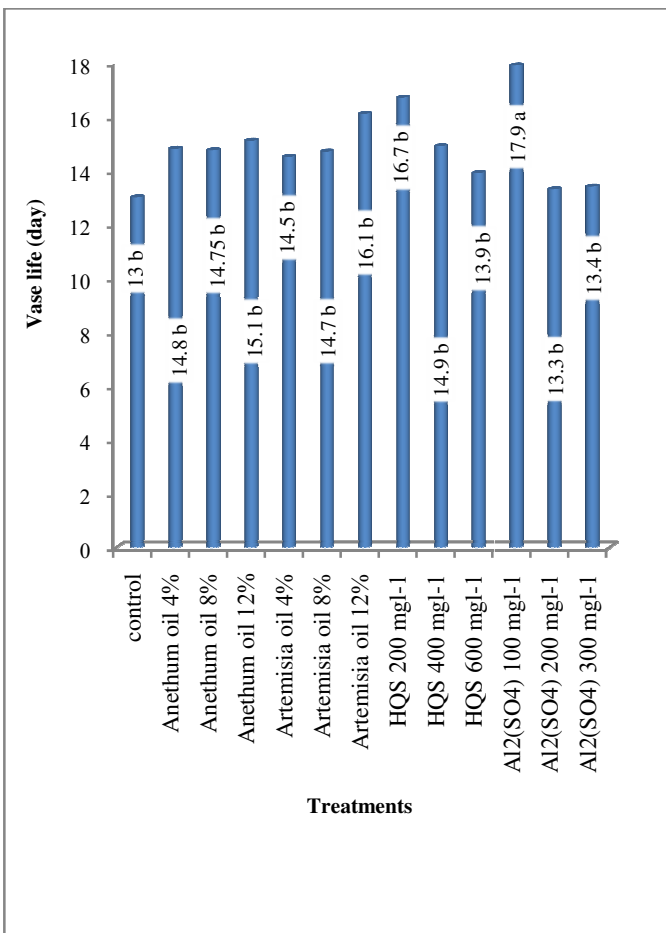


Figure 2. Effect of different treatments on vase life of cut carnation cv. 'Tempo'

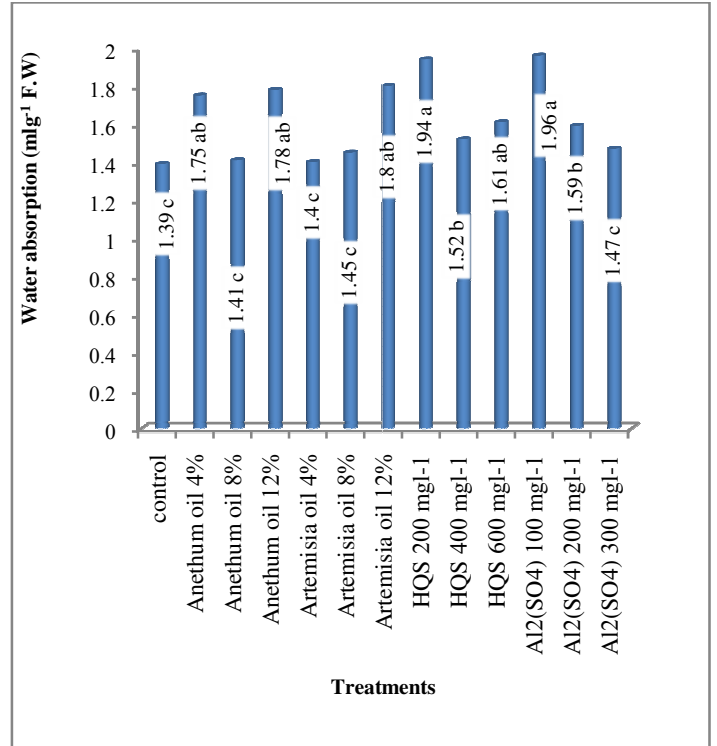


Figure 3. Effect of different treatments on water absorption of cut carnation cv. 'Tempo'

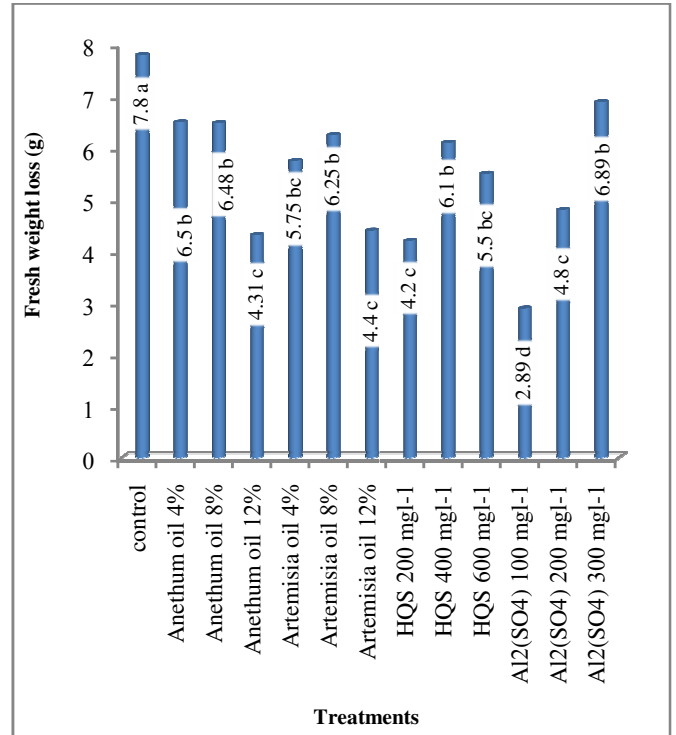


Figure 4. Effect of different treatments on fresh weight loss of cut carnation cv. 'Tempo'

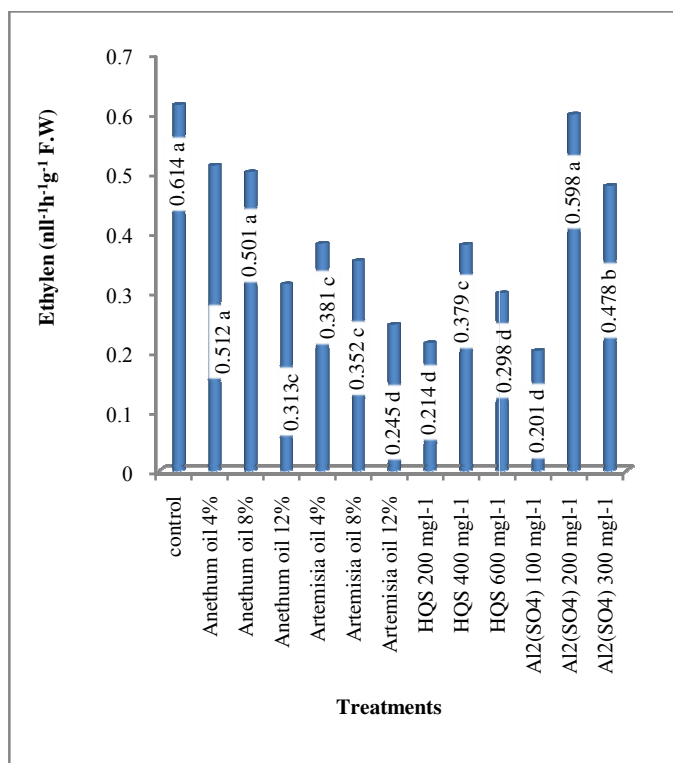


Figure 5. Effect of different treatments on ethylene production of cut carnation cv. 'Tempo'

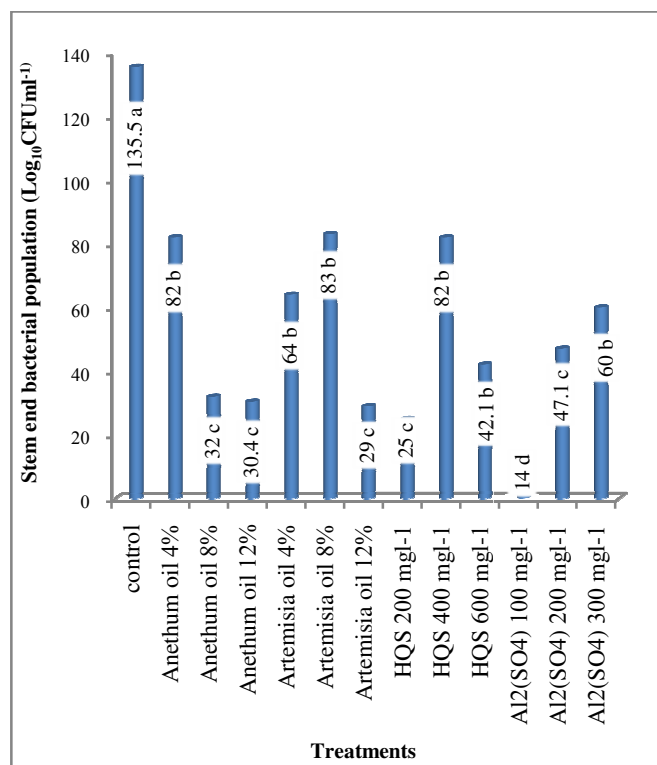


Figure 6. Effect of different treatments on stem end bacterial population of cut carnation cv. 'Tempo'

4. ACKNOWLEDGEMENTS

The authors would like to thank the Deputy of Research and Technology of Islamic Azad University of Rasht especially Dr. Ramin Shabani and Dr. Ali Mohammadi Torkashvand for their moral and financial supports.

REFERENCES

- [1] Abbaszadeh, M. and Elahian, H., "Investigation effect of some natural compounds on vase life of cut chrysanthemum cv. Mohandesi Zard", *Proceeding of The 7th International Horticultural Science Congress*, 2011, pp. 526-527 (In Farsi).
- [2] Anju, B., Tripathi, S. N., Sehgal, O. P., and Bhat, A., "Effect of pulsing, packaging and storage treatments on vase life of *Chrysanthemum* cut flowers", *Advances in Horticulture and Forestry*, 1999, 6: 125-131.
- [3] Arab, M., Khalighi, A., Arzani, K., and Naderi, R., "Investigation effect of cold storage, 8-hydroxyquinoline sulphate and sucrose on vase life and keeping quality of cut *Mattiola incana*", *Journal of Agricultural Science*, 2006, 37 (1): 83-92 (In Farsi).
- [4] Basiri, Y., and Zarei, H., "Effects of nano-silver treatments on longevity and some qualitative character of cut flowers of carnation (*Dianthus caryophyllus* cv. 'White Liberty')", *Proceeding of The 7th International Horticultural Science Congress*, 2011, pp. 2477-2478 (In Farsi).
- [5] Basiri, Y., Zarei, H., and Mashayekhi, K., "Effects of nano-silver treatments on vase life of cut flowers of carnation (*Dianthus caryophyllus* cv. 'White Liberty')", *Journal of Advanced Laboratory Research in Biology*, 2011, 2 (2): 49-55.
- [6] Bayat, H., Azizi, M., Shoor, M., and Vahdati, N., "Effect of ethanol and essential oil of medicinal plants on vase life of cut carnation cv. 'Yellow Candy'", *Journal of Horticultural Science*, 2011, 25 (4): 384-390 (In Farsi).
- [7] Burt, S., "Essential oils: Their antibacterial properties and potential applications in foods", *International Journal of Food Microbiology*, 2004, 94: 223-253.
- [8] Di, W., "Effects of antibiotics on the senescence of *Gerbera jamesonii* cut flower", *Journal of Anhui Agriculture Sciences* (Abstract), 2008.
- [9] Edrisi, B., "Postharvest physiology of cut flowers", *Payam-e-Digar Publication*, 2010, 150 p (In Farsi).
- [10] Edrisi, B., and Kalayie, A., "Investigation effect of chemical treatments on vase life and some quantitative character of cut rose flower", *Final Report to Agricultural Research Center and Natural Resource, Markazi Province* (In Farsi), 2005.
- [11] Elgimabi, M. N., and Ahmad, O. K., "Effects of bactericide and sucrose pulsing on vase life of rose cut flowers (*Rosa hybrida* L.)", *Botany Research International*, 2009, 2 (3): 164-168.
- [12] Esfandiari, B., Rezaei, A., Nemati, J., Tehranifar, A., and Ashrafi, J., "Effect of silver nanoparticles on postharvest life of lily cv. Shocking", *Proceeding of the 7th International Horticultural Science Congress*, 2011, pp. 2444-2446 (In Farsi).
- [13] Fahmy, A. R., and Hassan, S., "Postharvest studies on some important flower crops", www.lib.uni-corvinus.hu/Phd/Sadek-Hassan.2005.

- [14] Faraji, S., "Investigation effects of different chemical treatments on vase life of cut rose cv. Marosia", *M.Sc. Thesis, Olum va Tahghighat University, Tehran, Iran*, 2005, 102 p (In Farsi).
- [15] Farrager, R. S., Daw, Z. Y., and Abo-Raya, S., "Influence of some spice essential oils on *Aspergillus parasiticus* growth and production of aflatoxin in a synthetic medium", *Journal of Food Science*, 1989, 54: 74-76.
- [16] Figueroa, I., Colinas, M. T., Mejia, J., and Ramirez, F., "Postharvest physiological changes in rose of different vase life", *Cien Investigation of Agriculture*, 2005, 32: 167-176.
- [17] Goszcynska, D. M., and Rudnicki, R. M., "Storage of cut flower", *Horticulture Review*, 1981, 3: 59-146.
- [18] Hashemabadi, D., "Comparison effect of silver thiosulphate and silver nanoparticle on vase life of cut carnation cv 'Tempo'", *Final Report to Islamic Azad University, Rasht Branch, Iran*, 2012, 101 p (In Farsi).
- [19] Hashemabadi, D., Mostofi, Y., Kashi, A., Memariani, M., Shafiei, M., and Bagheri Moradi, G., "Investigation of the effect of 1-MCP (1-Methylcyclopropene) pretreatment in delaying of senescence of carnation (*Dianthus caryophyllus* L. 'Tempo')", *Journal of Horticultural Science and Technology*, 2007, 8: 13-20 (In Farsi).
- [20] Hojjati, Y., Khalighi, A., and Farrokhzad, A. R., "Chemical treatments of *Eustoma* cut flower cultivars for enhanced vase life", *Journal of Agriculture and Social Science*, 2007, 3 (3): 75-78.
- [21] Ichimura, K., Kojima, K., and Goto, R., "Effect of temperature, 8-hydroxyquinoline sulphate and sucrose on the vase life of cut roses flowers", *Postharvest Biology and Technology*, 1999, 15 (1): 33-40.
- [22] Jalili Marandi, R., Hassani, A., Abdollahi, A., and Hanafi, S., "Application of *Carum copticum* and *Saturega hortensis* essential oils and salicylic acid and silver thiosulphate in increasing the vase life if cut rose flowers", *Journal of Medicinal Plants Research*, 2011, 5 (20): 5034-5038.
- [23] Kazemi, M., and Ameri, A., "Response of vase life carnation cut flower to salicylic acid, silver nano-particles, glutamine and essential oil", *Asian Journal of Animal Science*, 2012, 6 (3): 122-131.
- [24] Kazemi, M., Hadavi, E., and Hekmati, J., "Role of salicylic acid in decrease of membrane senescence in cut carnation flowers", *Journal of Agricultural Technology*, 2011, 7 (5): 1417-14235.
- [25] Khalighi, A., "Floriculture", *Rozbahan Press*, 2010, 392 p (In Farsi).
- [26] Khan, F. U., Khan, F. A., Hayat, N., and Bhat, A., "Influence of certain chemicals on vase life of cut tulip", *Indian Journal of Plant Physiology*, 2007, 12: 127-132.
- [27] Kiamohammadi, M., "The effects of different floral preservative solutions on vase life of *Lisianthus* cut flowers", *Journal of Ornamental and Horticultural Plants*, 2011, 1 (2): 115-122.
- [28] Kim, Y., and Lee, J. S., "Changes in bent neck, water balance and vase life of cut rose cultivars as affected by preservative solution", *Journal of Korean Society for Horticultural Science*, 2002, 43 (2): 201-207.
- [29] Liao, L. J., Lin, Y. H., Huang, K. L., and Chen, W. S., "Vase life of *Eustoma grandiflorum* as affected by aluminum sulfate", *Botany Bulletin of Academic Science*, 2001, 42: 35-38.
- [30] Liu, J., He, S., Zhang, Z., Cao, J., Kv, P., He, S., Chen, G., and Joyce, D.C., "Nano-silver pulse treatments inhibit stem-end bacteria on cut gerbera cv. 'Ruikou' flowers", *Postharvest Biology and Technology*, 2009, 54: 59-62.
- [31] Lu, P., Cao, J., He, S., Liu, J., Li, H., Cheng, G., Ding, Y., and Joyce, D. C., "Nano-silver pulse treatments improve water relations of cut rosa cv. Movie Star flowers", *Postharvest Biology and Technology*, 2010, 57: 196-202.
- [32] Mousavi Bazaz, A., and Tehranifar, A., "Effects of ethanol, methanol and essential oils as novel agents to improve vase life of *Alstroemeria* flowers", *Journal of Biology and Environmental Science*, 2011, 5 (14): 41-46.
- [33] Nabigol, A., Naderi, R., Babalar, M., and Kafi, M., "Effects of some chemical treatment and cold storage on longevity of cut chrysanthemum", *Persian Paper Bank. Number of 511 Papers* (In Farsi), 2006.
- [34] Omidbeigi, R., "Production and processing of medicinal plants", *Astan Quds Razavi Publication* (In Farsi), 2004.
- [35] Oraee, T., Asgharzadeh, A., Kiani, M., and Oraee, A., "The role of preservative compounds on number of bacteria on the end of stems and vase solution of cut *Gerbera*", *Journal of Ornamental and Horticultural Plants*, 2011, 1 (3): 161-166.
- [36] Reid, M. S., Paul, J. L., Farhoomand, M. B., Kofranek, A. M., and Staby, G. L., "Pulse treatment with silver thiosulphate complex extends the vase life of cut carnation", *Journal of the American Society for Horticultural Science*, 1980., 105: 25-27.
- [37] Satoh, S., Shibuya, K., Waki, K., and Kosugi, Y., "Mechanism of senescence in carnation flowers", *Acta Horticulture*, 2005, 669: 191-198.
- [38] Shanani, N. T., "Application of essential oils to prolong the vase life of rose (*Rosa hybrida* L. cv. 'Grand') cut flowers", *Journal of Horticultural Science and Ornamental Plants*, 2012, 4 (1): 66-74.
- [39] Sharififar, F., Moshafi, M. H., Mansouri, S. H., Khodashenas, M., and Khoshnoodi, M., "In vitro evaluation of antibacterial and antioxidant activities of the essential oil and methanol extract of endemic (*Zataria multiflora*)", *Food Control*, 2007, 18: 800-805.
- [40] Solgi, M., Kafi, M., Taghavi, M., and Naderi, R., "Essential oils and silver nano particles (SNP) as novel agents to extend vase life of gerbera (*Gerbera jamesonii* cv. 'Dune') flowers", *Postharvest Biology and Technology*, 2009, 53: 155-158.
- [41] Svircev, A. M., Smith, R. J., Zhou, T., Hernade, M., Liu, W., and Chid, C. L., "Effects of thymol fumigation on survival and ultrastructure of *Monilinia fructicola*", *Postharvest Biology and Technology*, 2007, 45: 228-233.
- [42] Tar, T., and Hassan, F. A., "Evaluating vase life and tissue structure of some compositae (Asteraceae) species", *Internal Journal of Horticultural Science*, 2003, 9: 87-89.
- [43] Van Doorn, W. G., "Water relation of cut flowers", *Horticulture Review*, 1997, 18: 1-85.